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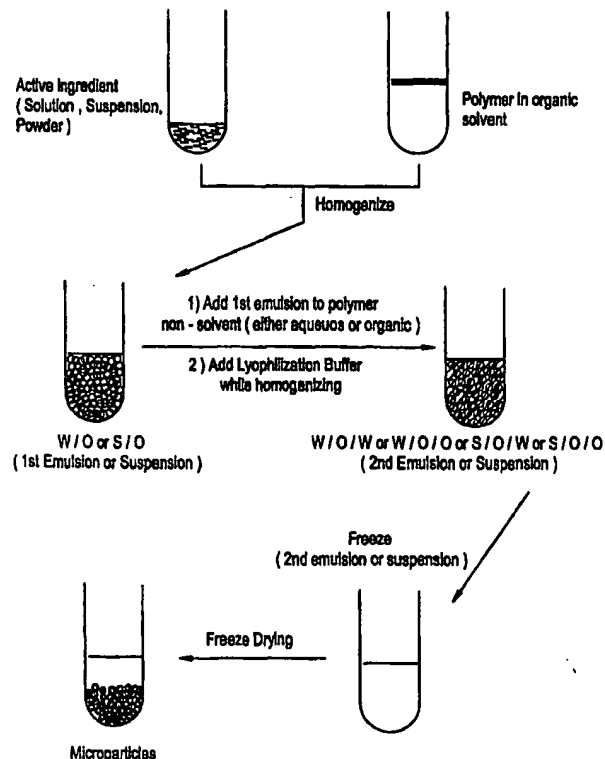
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(54) Title: BIODEGRADABLE MICROPARTICLES FOR THE SUSTAINED DELIVERY OF THERAPEUTIC DRUGS

(57) Abstract

The present invention relates to improved methods of making polymeric microparticles containing a variety of active ingredients, e.g. protein drugs. In addition, the present invention relates to using the above active protein containing polymeric microparticles to prepare compositions for the sustained delivery of the therapeutics.



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BIODEGRADABLE MICROPARTICLES FOR THE SUSTAINED DELIVERY
OF THERAPEUTIC DRUGS

This application is a continuation-in-part,
of application Serial No. 08/843,975 filed April 17,
1997 which is hereby incorporated by reference.

FIELD OF THE INVENTION

The present invention generally relates to
improved methods of making biodegradable polymeric
microparticles containing an active ingredient. In
addition, the present invention relates to use of said
microparticles to prepare compositions for the
sustained delivery of therapeutics.

BACKGROUND OF THE INVENTION

Due to recent advances in genetic and cell
engineering technologies, proteins known to exhibit
various pharmacological actions *in vivo* are capable of
production in large amounts for pharmaceutical
applications. Such proteins include erythropoietin
(EPO), granulocyte colony-stimulating factor (G-CSF),
interferons (alpha, beta, gamma, consensus), tumor
necrosis factor binding protein (TNFbp), interleukin-1
receptor antagonist (IL-1ra), brain-derived
neurotrophic factor (BDNF), keratinocyte growth factor
(KGF), stem cell factor (SCF), megakaryocyte growth
differentiation factor (MGDF), osteoprotegerin (OPG),
glial cell line derived neurotrophic factor (GDNF) and

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obesity protein (OB protein). OB protein may also be referred to herein as leptin.

Because these proteins generally have short in vivo half-lives and negligible oral bioavailability, they are typically administered by frequent injection, thus posing a significant physical burden on the patient and associated administrative costs. As such, there is currently a great deal of interest in developing and evaluating sustained-release formulations. Effective sustained-release formulations can provide a means of controlling blood levels of the active ingredient, and also provide greater efficacy, safety, patient convenience and patient compliance. Unfortunately, the instability of most proteins (e.g. denaturation and loss of bioactivity upon exposure to heat, organic solvents, etc.) has greatly limited the development and evaluation of sustained-release formulations.

Attempts to develop sustained-release formulations have included the use of a variety of biodegradable and non-biodegradable polymer (e.g. poly(lactide-co-glycolide)) microparticles containing the active ingredient (see e.g., Wise et al., *Contraception*, 8:227-234 (1973); and Hutchinson et al., *Biochem. Soc. Trans.*, 13:520-523 (1985)), and a variety of techniques are known by which active agents, e.g. proteins, can be incorporated into polymeric microspheres (see e.g., U.S. Patent No. 4,675,189 and references cited therein).

One such technique is spray-drying, wherein the polymer and active ingredient are mixed together in a solvent for the polymer, and then the solvent is evaporated by spraying the solution, leaving polymeric droplets containing the active ingredient. For a

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detailed review of spray drying see e.g. Masters, K.,
"Spray Drying Handbook" (John Wiley & Sons, eds., New
York 1984). Although the spray drying technique has
proven useful in certain instances, it still suffers
5 from the fact that biologically active proteins are
often denatured due to contact with the organic polymer
and solvent, or due to the heat generated during the
spray drying processes.

Another technique which can be used to form
10 microspheres is solvent evaporation. Solvent
evaporation involves the dissolving of the polymer in
an organic solvent which contains either dissolved or
dispersed active ingredient. The polymer/active
ingredient mixture is then added to an agitated
15 continuous phase which is typically aqueous.
Emulsifiers are included in the aqueous phase to
stabilize the oil-in-water emulsion. The organic
solvent is then evaporated over a period of several
hours or more, thereby depositing the polymer around
20 the core material. For a complete review of the
solvent evaporation procedure see e.g. U.S. Patent No.
4,389,330 (and references cited therein). As with the
spray drying technique, solvent evaporation techniques
have proven useful in certain instances. However, the
25 technique is often not preferred because active
ingredient is often lost during the solvent extraction
process. This is because the process involves
emulsification into an aqueous phase, and a water
soluble drug will often rapidly partition from the more
30 hydrophobic polymer-solution phase into the aqueous
surroundings.

Yet another technique which can be used to
form microspheres is phase separation, which involves
the formation of a water-in-oil emulsion or oil in

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water emulsion. The polymer is precipitated from the continuous phase onto the active agent by a change in temperature, pH, ionic strength or the addition of precipitants. For a review of phase separation techniques, see e.g. U.S. Patent No. 4,675,800 (and references cited therein). Again, this process suffers primarily from loss of active ingredient due to denaturation.

The release characteristics for the active ingredient from microparticles prepared by methods such as those described above may be continuous or discontinuous, and in some cases, the initial level of active ingredient release is too high or too low. Thus, various additives are often utilized in an attempt to control the release of active ingredient (see e.g., EP 0 761 211 A1, published March 12, 1997).

To avoid the denaturation of protein and other fragile biological molecules which occurs upon spray drying, solvent evaporation or phase separation by classical techniques, the emulsion of polymers and active ingredient can be atomized into frozen nonsolvent overlayed with liquified gas such as nitrogen to form particles, and then extracted at very low temperatures. The extremely low processing temperatures may preserve the activity and integrity of the fragile biological molecules such as proteins. However, the method leads to poor loading efficiencies and yields, resulting in the loss of precious biological material, and is cumbersome, difficult and expensive to implement at the large scales required for commercial production.

Clearly the need still exists for an improved method for preparing polymeric microparticles containing an active ingredient which is simple,

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inexpensive, versatile, and, most importantly, which protects against loss of protein activity and which provides for high loading efficiencies and yields, thereby allowing for more consistent active ingredient
5 release over an extended period of time.

SUMMARY OF THE INVENTION

10 As fully described below, the present invention provides an improved method for preparing polymeric microparticles containing an active ingredient through unique utilization of direct lyophilization of emulsion or suspension. This
15 improved method provides several significant advantages over the processes described in the art, including, for example, 1) ease of manufacture of the active ingredient loaded microparticles (i.e. fewer and less cumbersome steps); and 2) the provision of sustained
20 release formulations that maintain the activity and integrity of the active ingredient during release, thus providing for controlled release of active ingredient over an extended period of time. Additionally, the processes of the present invention provide the
25 advantages of versatility as relates to the class of polymers and/or active ingredient which may be utilized, as well as attainment of higher yields, high loading, and higher loading efficiencies.

Accordingly, one aspect of the present
30 invention relates to a new and improved process for preparing a composition comprising an active ingredient contained within polymeric microparticles, wherein a mixture of the active ingredient and the polymer are dispersed within a continuous phase, the resulting

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dispersion is frozen, and the water and organic solvents removed from the dispersion by lyophilization. Importantly, the present process is more refined and simpler than those described in the art, and the activity and integrity of the active ingredient is maintained throughout the process.

The present process can be generally described as comprising the steps of: (a) preparing a polymeric solution; (b) adding an active ingredient to produce a mixture; (c) dispersing said mixture within a continuous phase, i.e. non-solvent(s), to produce a dispersion; (d) adding an excipient to said dispersion; (e) freezing said dispersion to produce a frozen mixture; and (f) lyophilizing said frozen mixture to produce the desired active ingredient containing microparticles. Alternatively, step (b) can be omitted and "blank" microparticles prepared, onto which active ingredient is then loaded by suspending the blank microparticles in active ingredient solution.

A second aspect of the present invention is a pharmaceutical composition for the sustained-release of an active ingredient comprising a biologically active ingredient contained within polymeric microparticles, or, alternatively, a biologically active ingredient loaded onto the polymeric microparticles. Importantly, the sustained-release compositions of the present invention maintain the activity and integrity of the active ingredient during encapsulation and release, which helps to provide for longer periods of consistent release.

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BRIEF DESCRIPTION OF THE FIGURES

Figure 1 is a schematic of the process of the present invention for making the active ingredient containing microparticles.

Figure 2 is a plot depicting the *in vitro* release of various protein loaded microparticles. Microparticles containing leptin (incorporated in liquid form) is depicted by the -▲- line; microparticles containing Zn:leptin (incorporated in suspension form) is depicted by the -●- line; and microparticles containing Zn:leptin (incorporated in powder form) is depicted by the -■- line. % leptin released (leptin concentration having been determined by UV spectrophotometer at 280nm) is plotted vs. time (days).

Figure 3 shows the circular dichroism (CD) data comparing leptin released from leptin loaded microparticles (*in vitro*) on day 7 (solid line) vs. a control sample of leptin in formulation buffer (dashed line). CD spectra were obtained with a Jasco J-720 spectropolarimeter (Japan Spectroscopic Co., Tokyo, Japan). The samples (3.5 μ M as determined by A_{280}) were analyzed at 22°C with a cell path length of 0.1 cm.

Figure 4 shows the high performance liquid chromatography (HPLC) data for leptin released from leptin loaded microparticles (*in vitro*) at various time points (0 to 168 hours). Analytical Size Exclusion Chromatography (SEC) was performed with a TosoHaas G2000 SW column (Montgomery, PA) using a Waters HPLC

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(Milford, MA) with 20mM sodium phosphate, 125mM NaCl, pH 7.4 at 0.8 mL/min. Absorbance at 280nm is plotted vs. run time (minutes).

5 Figure 5 is a picture of an SDS-PAGE gel
(2-20% Tris-glycine gel (Novex, San Diego, CA))
containing the following samples: lane 1: leptin
standard; lane 2: molecular weight standards;
lanes 3-9: leptin released from leptin loaded
10 microparticles (in vitro) at 2 hours, 24 hours,
68 hours, 92 hours, 116 hours, 140 hours, and
168 hours, respectively; lane 10, molecular weight
standards. Samples were diluted with nonreducing SDS
buffer, heated at 100°C for five minutes and 1 µg of
15 protein loaded into each well. The gels were stained
with Coomassie Blue R-250.

Figure 6 shows *in vivo* bioactivity of leptin
loaded microparticles in normal mice, in terms of
20 % body weight loss relative to buffer control for a
7 day period. Buffer control (-*-) is plotted vs.
leptin injected @ 10 mg/kg daily (-●-) vs. leptin
injected @ 50 mg/kg on day 0 only (-x-) vs. leptin
loaded microparticles injected @ 50 mg/kg on day 0 only
25 (-■-) vs. control microparticles injected on day 0 only
(-○-).

Figure 7 is a plot depicting the *in vitro*
release of BDNF from microparticles onto which BDNF had
30 been absorbed. % BDNF released (BDNF protein
concentration having been determined by UV
spectrophotometer at 280nm) is plotted vs. time (days).

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DETAILED DESCRIPTION

Polymers may be selected from the group consisting of biocompatible and/or biodegradable polymer. As defined herein, biodegradable means that the composition will erode or degrade *in vivo* to form smaller chemical species. Degradation may occur, for example, by enzymatic, chemical or physical processes. Suitable biodegradable polymers contemplated for use in the present invention include poly(lactide)s, poly(glycolide)s, poly(lactic acid)s, poly(glycolic acid)s, polyanhydrides, polyorthoesters, polyetheresters, polycaprolactone, polyesteramides, polycarbonate, polycyanoacrylate, polyurethanes, polyacrylate, blends and copolymers thereof.

The range of molecular weights contemplated for the polymers to be used in the present processes can be readily determined by a person skilled in the art based upon such factors the desired polymer degradation rate. Typically, the range of molecular weight will be 2000 to 2,000,000 Daltons. Almost any type of polymer can be used provided the appropriate solvent and non-solvent are found.

The term "PLGA" as used herein is intended to refer to a polymer of lactic acid alone, a polymer of glycolic acid alone, a mixture of such polymers, a copolymer of glycolic acid and lactic acid, a mixture of such copolymers, or a mixture of such polymers and copolymers. Preferably, the biodegradable polymer will be poly lactide-co-glycolide (PLGA).

Unless otherwise noted, the term microparticles can be used to encompass microparticles, microspheres, and microcapsules. Active agents to be incorporated into the microparticles are synthetic or

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natural compounds which demonstrate a biological effect when introduced into a living creature. Contemplated active agents include peptides, small molecules, carbohydrates, nucleic acids, lipids, and proteins.

5 Proteins contemplated for use include potent cytokines, including various hematopoietic factors such as granulocyte-colony stimulating factors (see, U.S. Patent Nos. 4,810,643, 4,999,291, 5,581,476, 5,582,823, and PCT Publication No. 94/17185, hereby incorporated

10 by reference including drawings), GM-CSF, M-CSF, MGDF, the interferons (alpha, beta, gamma, omega), interferon consensus (see, U.S. Patent Nos. 5,372,808, 5,541,293 4,897,471, and 4,695,623 hereby incorporated by reference including drawings), the interleukins (1-12)

15 (see, U.S. Patent No. 5,075,222, hereby incorporated by reference including drawings), erythropoietin (EPO) (see, U.S. Patent Nos. 4,703,008, 5,441,868, 5,618,698 5,547,933, and 5,621,080 hereby incorporated by reference including drawings), fibroblast growth

20 factor, TNF, TNFbp, IL-1ra, stem cell factor (PCT Publication Nos. 91/05795, 92/17505 and 95/17206, hereby incorporated by reference including drawings), nerve growth factor, GDNF, BDNF, NT3, platelet-derived growth factor, and tumor growth factor (alpha, beta),

25 osteoprotegerin (OPG), and OB protein (leptin).

Also contemplated for incorporation into the compositions of the present invention are derivatives, fusion proteins, conjugates, analogs or modified forms of the natural active ingredients. Chemical

30 modification of biologically active proteins has been found to provide additional advantages under certain circumstances, such as increasing the stability and circulation time of the therapeutic protein and decreasing immunogenicity. For example, U.S. Patent

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No. 4,179,337, Davis et al., issued December 18, 1979, discloses conjugation of water-soluble polypeptides such as enzymes and insulin to polyethylene glycol (PEG); see also WO 87/00056, published January 15, 5 1987.

Another type of chemical modification contemplated for the active ingredients of the present invention is succinylation. The properties of various succinylated proteins are described in Holcenberg et 10 al., *J. Biol. Chem*, 250:4165-4170 (1975), and WO 88/01511 (and references cited therein), published March 10, 1988.

The present leptins used are preferably those with amino acid sequence of natural human OB protein; 15 see Zhang et al., *Nature* 372:425-432 (1994); see also, the Correction at *Nature* 374: 479 (1995), optionally with an N-terminal methionyl residue incident to bacterial expression is used. (See, Materials and Methods, *infra*). PCT publication No. WO 96/05309, 20 published February 22, 1996, entitled, "Modulators of Body Weight, Corresponding Nucleic Acids and Proteins, and Diagnostic and Therapeutic Uses Thereof" fully sets forth OB protein and related compositions and methods, and is herein incorporated by reference. An amino acid 25 sequence for human OB protein is set forth at WO 96/05309 Seq. ID Nos. 4 and 6 (at pages 172 and 174 of that publication), and the first amino acid residue of the mature protein is at position 22 and is a valine residue. The mature protein is 146 residues (or 145 if 30 the glutamine at position 49 is absent, Seq. ID No. 4). Specific leptin derivatives contemplated for use in the present invention include Fc-leptin fusions, succinylated-leptin, and zinc derivatized leptin (Zn:leptin). It is desirable to have such leptin

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containing sustained-release compositions as such compositions could serve to enhance the effectiveness of either exogenously administered or endogenous leptin, or could be used, for example, to reduce or
5 eliminate the need for exogenous leptin administration.

In general, an aqueous solution, a suspension, or a solid form of the active agent can be admixed with the organic solvent containing the polymer. When an aqueous solution of active ingredient
10 is used, polymer:active ingredient emulsions will be formed and used to prepare microparticles. When a suspension or solid form of active ingredient is used, polymer:active ingredient suspensions are formed and used to prepare the microparticles.

15 The principal embodiment of the method for making the protein loaded microparticles comprises:
(a) dissolving a polymer in an organic solvent to produce a polymeric solution; (b) adding an active ingredient in a form selected from the group consisting
20 of an aqueous solution, a suspension, and a powder to said polymeric solution to produce a active ingredient-polymer mixture comprising a first emulsion or suspension; (c) dispersing said first emulsion or suspension within a continuous phase, i.e., non-
25 solvent(s), to produce a dispersion; (d) adding an excipient to said dispersion to produce a final dispersion; (e) freezing said final dispersion; and (f) lyophilizing said frozen final dispersion to remove different solvents (aqueous and organic) to produce the
30 desired protein loaded microparticles. The process is shown schematically in Figure 1. As depicted in Figure 1, step c) can alternatively comprise diluting said first emulsion or suspension with a polymer non-solvent. Additionally, step a) can comprise active

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ingredient being dissolved directly in the organic polymer solution to form a homogeneous first mixture.

The solvent to be used for dissolving the PLGA in step a) of the present processes includes, for example, chloroform, ethyl acetate, methylene chloride, acetonitrile, THF and acetone. In one embodiment of the present invention, the solvent to be used is chloroform. Non-solvents contemplated for use in step c) include water, hexane, ethanol, methanol, and carbon tetrachloride or mixtures thereof (e.g. water/ethanol).

The polymer concentrations contemplated for use in the processes of the present invention are in the range of 5-70 gm/100mL. In the embodiments of the present invention which utilize PLGA, the polymer concentration will preferably be in the range of 10-20 gm/100mL.

The protein concentrations contemplated for use in the processes of the present invention are in the range of 0-300 mg/mL when in solution or suspension, or equivalent solid protein. In the embodiments of the present invention which utilize leptin, the protein concentration is preferably 100 mg/mL.

For the emulsions produced in the processes of the present invention, the organic:aqueous ratios contemplated for use are 1:1 to 12:1. In the embodiments of the present invention which utilize PLGA and leptin, the organic:aqueous ratio will preferably be 4:1 for the first emulsion. In general, the microparticles prepared by the methods of the present invention will generally comprise 0-60% by weight of protein.

The addition of a lyophilization excipient in step d) of the process described above was found to be

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useful to insure that the microparticles did not aggregate or fuse during lyophilization. One or more excipients may be added. Importantly, such excipient(s) could also be added in step b) or c) of the process. The lyophilization excipient(s) contemplated for use in the present processes include lactose, mannitol, dextran, sucrose, heparin, glycine, glucose, glutamic acid, gelatin, sorbitol, dextrose, trehalose, methocel, hydroxy ethyl cellulose, hydroxy ethyl starch, poly(ethylene glycol), poly(vinyl pyrrolidone) and polyvinyl alcohol, or various combinations thereof, as well as other buffers, protein stabilizers, cryoprotectants, and cryopreservatives commonly used by those skilled in the art.

The temperatures contemplated for use in the freezing step (step e) of the present processes are in the range of -283°C (liquid nitrogen) to -20°C. These temperatures are utilized so as to stabilize the emulsions or suspensions. The final emulsion or suspension can be frozen immediately using the temperatures described above, or can be stored at room temperature prior to freezing. In one embodiment of the present invention, the final emulsion was frozen immediately and the temperature utilized for the freezing was -80°C.

The temperatures contemplated for use in step f) of the present processes are in the range of -100°C to room temperature. Preferably, the temperature of the frozen sample of step e) will be lowered -80°C and held for one hour prior to being connected to the vacuum system. The temperature is then raised step-wise in 5°C/hour increments to -25°C to effect removal of the aqueous phase and any residual organic phase. The sample is then held under vacuum

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for 4-5 days (or whenever the vacuum gauge indicates no more vapor removal), and then the temperature raised to -5°C for 6-8 hours before the removing the sample from the vacuum system. It is utilization of this single
5 step, i.e., direct lyophilization of the final emulsion or suspension, which refines and simplifies the present process over previously described processes, which require multiple steps and are often cumbersome. And, importantly, this direct lyophilization provides the
10 enhanced stability for a wide variety of active ingredient *in vivo*, as well as the attainment of higher loading, higher loading efficiencies, and higher yields. Thus, the significant advantages of the present processes as compared to the processes
15 described in the art, include for example, 1) ease of manufacture of the active ingredient loaded microparticles; 2) versatility as relates to the class of polymers and/or active ingredients which may be utilized; 3) higher yields and loading efficiencies;
20 and 4) the provision of sustained release formulations that release active, intact active ingredient *in vivo*, thus providing for controlled release of active ingredient over an extended period of time (e.g. up to 180 days). As used herein the phrase "contained
25 within" denotes a method for formulating an active ingredient into a composition useful for controlled release, over an extended period of time of the active ingredient.

In the sustained-release compositions of the
30 present invention, an effective amount of active ingredient will be utilized. As used herein, sustained release refers to the gradual release of active ingredient from the polymer matrix, over an extended period of time. The sustained release can be

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continuous or discontinuous, linear or non-linear, and this can be accomplished using one or more polymer compositions, drug loadings, selection of excipients, or other modifications.

5 In general, comprehended by the present invention are pharmaceutical compositions comprising effective amounts of protein or derivative products of the invention together with pharmaceutically acceptable diluents, stabilizers, preservatives, solubilizers, emulsifiers, adjuvants and/or carriers. Such
10 compositions include diluents of various buffer content (e.g., Tris-HCl, phosphate), pH and ionic strength; additives such as detergents and solubilizing agents (e.g., Tween 80, Polysorbate 80), anti-oxidants (e.g.,
15 ascorbic acid, sodium metabisulfite), preservatives (e.g., Thimersol, benzyl alcohol) and bulking substances (e.g., lactose, mannitol); see, e.g., Remington's Pharmaceutical Sciences, 18th Ed. (1990, Mack Publishing Co., Easton, PA 18042) pages 1435-1712
20 which are herein incorporated by reference. An effective amount of active ingredient is a therapeutically, prophylactically, or diagnostically effective amount, which can be readily determined by a person skilled in the art by taking into consideration
25 such factors as body weight, age, therapeutic or prophylactic or diagnostic goal, and release rate desired.

 A suspension of protein loaded microparticles prepared in accordance with the present invention is
30 preferably administered by injection intraperitoneally, subcutaneously, or intramuscularly. However, it would be clear to one skilled in the art that other routes of delivery could also be effectively utilized using the compositions of the present invention.

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The following examples are offered to more fully illustrate the invention, but are not to be construed as limiting the scope thereof. Example 1 describes the novel method for preparing protein loaded microparticles. Leptin (in the form of an aqueous solution) is used as an example protein and the ability of leptin loaded microparticles to provide for sustained release of leptin, both *in vitro* and *in vivo* is demonstrated. Example 2 demonstrates that different polymers can be used to make leptin loaded microparticles. Example 3 demonstrates that different leptin derivatives, as well as entirely different proteins (all in the form of an aqueous solution), can be used in the novel methods of the present invention. Example 4 demonstrates the effects of temperature on the freezing step and on the lyophilization step of the process of the present invention. Example 5 demonstrates that different organic solvents can be used to dissolve the PLGA polymers in the methods of the present invention. Example 6 demonstrates that a Zn:leptin suspension and Zn:leptin lyophilized powder can be used in the novel methods of the present invention. Example 7 demonstrates that a spray-dried protein, spray-dried IL-1ra, can be used in the novel methods of the present invention. Example 8 demonstrates that "blank" microparticles onto which active ingredient (e.g. BDNF) has been absorbed, can also provide for sustained release of active ingredient *in vitro*. Materials and methods follow.

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EXAMPLE 1

5 This example describes the novel methods for preparing protein loaded microparticles; specifically, the preparation of poly(D,L-lactide-co-glycolide) microspheres containing leptin.

0.6 g of RG-502H, poly(D,L-lactide-co-glycolide) (Boehringer Ingelheim Chemicals (B.I. Chemicals), Henley Div., Montvale, NJ) was dissolved in 10 4 mL of chloroform and filtered through a 0.2 μ m PTFE filter. 1 mL of leptin at 100 mg/mL in 10mM sodium acetate, pH 4.8 (prepared as described in Materials and Methods, *infra*), was first sterile filtered and then 15 gently added to the top of the polymer solution. The two layers were homogenized using a Polytron homogenizer (PT-DA3012/2T generator, Brinkman, Westbury, NY) at 15,000 to 20,000 rpm for 30-45 seconds while the emulsion container was immersed in an ice 20 bath.

The resultant first emulsion (w/o) was added to 10 mL of water while homogenizing at 15,000 rpm for 20-30 sec. To the resulting second emulsion (w/o/w), 1 mL of lyophilization excipient (100 mg/mL Glycine, 25 100 mg/mL Sucrose, 10 mg/mL polyvinyl-alcohol (PVA) [22,000 M.W., 88% Hydrolyzed], 10% v/v ethanol) was added and briefly homogenized to insure thorough mixing. The final emulsion under optical microscope showed 1-10 μ m free flowing spheres. The final 30 emulsion was poured into a flask and frozen at -45°C.

The temperature of the bath was then reduced in one hour to -80°C. After an hour at -80°C, the flask was connected to a vacuum system and lyophilization first carried out at -80°C. The vacuum

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level was monitored so that removal of organic solvent could be determined by a drop in vacuum to the system level. The temperature was then raised step-wise in 5°C/hour increments to -25°C to effect the removal of
5 the aqueous phase and any residual organic phase.

After 4-5 days when the vacuum gauge indicated no more vapor removal, the temperature of the bath was raised to -5°C for 6-8 hours before removing the samples from the vacuum system. The microparticles
10 were weighed and then stored at -20°C until needed.

The process described above was also tested with the following modifications: 1) the first emulsion (w/o) or suspension (s/o) was added to 20 mL of water/ethanol (75%/25%) while homogenizing at 5,000 rpm
15 for 30 seconds; 2) 10-40 mL of either water or cold ethanol was added slowly to the second emulsion ((w/o/w) or (s/o/w)) and then the final emulsion incubated at room temperature for up to three hours prior to freezing; and 3) the final emulsion was
20 hardened by incubating it room temperature for different time intervals (0-4 hours) prior to freezing. In each instance, leptin loaded microparticles were obtained.

25 In vitro Release of Leptin from PLGA microparticles

"In vitro" release kinetics of leptin from the microparticles prepared as described above were determined by making a 20 mg/mL suspension of the
30 particles in 20mM sodium phosphate, 5% Sorbitol, pH 7.4 (alternatively, 20mM histidine could be used instead of phosphate). At each time interval, the microsphere suspension was centrifuged and leptin concentration in the supernatant was determined by UV

- 20 -

spectrophotometer at 280nm as well as by SEC-HPLC at 220nm. The % leptin released over time is depicted in Figure 2. The integrity of the leptin released from the PLGA microparticles was confirmed by circular dichroism (CD) (Figure 3), HPLC (Figure 4), *in vitro* bioassay and gel electrophoresis (SDS-PAGE) (Figure 5). The CD data showed retention of secondary structure, and HPLC and gel electrophoresis showed no obvious chemical degradation or aggregation.

10

In vivo Bioactivity of Leptin loaded microparticles

"*In vivo*" bioactivity of leptin loaded microparticles were evaluated in normal mice and rats by suspending 80-100 mg/mL microparticles in 20mM sodium phosphate, 5% Sorbitol, pH 7.4, buffer. The suspensions were prepared an hour before subcutaneous injection by placing the microparticles and buffer on a shaker in a 5°C cold room. All subsequent manipulations immediately prior to injection were done with refrigerated syringes and 25-gauge needles. % body weight loss relative to buffer control was determined for a 7 day period. After day 7, animals were sacrificed for histological examination of the injection site. A single injection of leptin loaded microparticles resulted in sustained weight loss in mice for 7 days period (Figure 6). Histological examination of the injection site revealed a localized minimal to mild inflammatory reaction, which was fully reversible with biodegradation of the microparticles over time.

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EXAMPLE 2

This example was designed to test the effectiveness of different molecular weights of PLGA or blends in the preparation of leptin loaded microparticles. The preparation and evaluation procedures described in Example 1 were utilized to test the various polymers listed in Table 2 below.

10

Table 2

	<u>Polymers:</u>	<u>Source:</u>
	RG-501H	B.I. Chemicals
	RG-502H	B.I. Chemicals
15	RG-502	B.I. Chemicals
	RG-503H	B.I. Chemicals
	(RG-501H):(RG-502H) blends*	B.I. Chemicals
	(RG-501H):(PEG/PLGA)**	B.I. Chemicals
20	* Polymer blends made by mixing 20:80 & 50:50 weight ratios of RG-501H and RG-502H	
	** 80:20 (by weight) blend of PLGA (501H) and PLGA(501H):PEG(1000) AB block copolymer.	

25

Using each of the polymers listed in Table 2, protein loaded microparticles could be effectively prepared.

30

EXAMPLE 3

This example describes the preparation of microparticles containing leptin derivatives as well as other proteins. The preparation and evaluation procedures described in Example 1 were utilized to test the various proteins listed in Table 3 below.

35

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Table 3

	Protein	Molecular Weight (Daltons)	Concentration: (mg/mL)	Formulation:
5	Leptin	16,158	50-130	10mM NaAcO, pH 4.8-8.0
			80	10mM NaAcO pH 4.8
10			100	+ 10% Sucrose
			60	Lyo Buffer ^a
				Lipid Complexed ^b
	20kd PEG-leptin	~36,158	64-122	10mM NaAcO, pH 4.8-8.0
15	Succinylated leptin	16,258	60-80	10mM NaPhos, pH 7.0-8.0
20	G-CSF	18,798	50-100	10mM NaAcO, pH 4.8-8.0
			60-100	10mM NaAcO, pH 4.8-8.0
			55	+ 5-16% Sucrose
			60	1mM NaCl, 10% Trehalose, pH 7.6
25	BDNF	13,513	45-120	Lipid Complexed ^b
	IL-1ra	17,258	100-200	100mM NaPhos, pH 7
30	TNFbp	18,278	105	10mM NaCitrate, 140mM NaCl, pH 6.5
35	BSA	66,262	100	10mM NaPhos, 2% Gly, 1% Sucrose, pH 7
				10mM NaAcO, pH 4.8

^a Lyophilization Buffer = 10 mg/mL Glycine, 5 mg/mL Sucrose, 10mM glutamic acid, pH 4.5.

40 ^b Lipid complexed 30:1 mol. ratio DMPG or DCPG to protein in 20mM NaAcO, pH 4.8.

With each of the proteins listed above, protein loaded microparticles were obtained, thus demonstrating the flexibility of the novel process of the present invention. And importantly, it is demonstrated that protein loaded microparticles can also be effectively prepared using different leptin derivatives.

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EXAMPLE 4

In this example, the effects of temperature
5 on the freezing step and lyophilization step of the
process of the present invention were evaluated. As
relates to the freezing step (step 5), liquid nitrogen,
-80°C and -45°C were tested. As relates to the
lyophilization step (step 6), -80°C, -45°C, and -25°C
10 were tested. The procedure described in Example 1 was
used to test the various temperatures and it was
determined that the tested temperatures had very little
effect on either step in the process.

15

EXAMPLE 5

In this example, different organic solvents
were tested to dissolve the PLGA polymers in the
20 methods of the present invention. The procedure
described in Example 1 was repeated using ethyl
acetate, methylene chloride. Each of the tested
organic solvents were found to be effective in the
methods of the present invention.

25

EXAMPLE 6

This example tested the ability of an active
30 ingredient suspension and/or lyophilized powder to be
used in the methods of the present invention.

A 100 mg/mL, in 10mM Tris, 50 µM Zinc
chloride, pH 7.0, Zn/OB protein suspension (prepared as
described in the Materials and Methods section below)

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was tested and evaluated as described in Example 1.

Also tested and evaluated was a 100 mg Zn:leptin lyophilized powder (prepared as described in the Materials and Methods section below). It was

5 demonstrated that the incorporated Zn:leptin suspension and Zn:leptin powder could be effectively utilized to prepare microparticles according to the novel methods of the present invention (see Figure 2 for *in vitro* release data).

10

EXAMPLE 7

This example tested the ability of a spray-dried protein, spray-dried IL-1ra, to be used in the
15 methods of the present invention. A 150 mg spray-dried IL-1ra powder (prepared as described in the Materials and Methods section below) was tested and evaluated as described in Example 1. It was demonstrated that the
20 spray-dried IL-1ra preparation could be effectively utilized to prepare microparticles according to the novel methods of the present invention

25

EXAMPLE 8

In this example, the method described in Example 1 was modified such that 20mM NaAcO, pH 4.8, was mixed initially with the polymer, resulting in the
30 preparation of "blank" microparticles. 6 mg of blank microparticles was then diluted with 1 mL of BDNF (4.4 mg/mL in 0.1M sodium phosphate, pH 6.9) and the mixture incubated at 37°C with shaking. After 2 hours, microparticles were isolated by centrifugation and the

- 25 -

unbound protein fraction was determined by UV spectrophotometer. 1.76 mg of BDNF was bound to polymer to give 22% protein loading on the microparticles. In vitro release kinetics were then
5 determined as described in Example 1. The % BDNF released over time is depicted in Figure 7.

Materials and Methods

10

1. Preparation of recombinant methionyl human leptin.

The present recombinant methionyl-human-leptin may be prepared according to the above
15 incorporated-by-reference PCT publication, WO 96/05309 at pages 151-159. For the present working examples, a human leptin was used which has (as compared to the amino acid sequence at page 158) a lysine at position 35 instead of an arginine, and an isoleucine at
20 position 74 instead of an isoleucine. Other recombinant human leptins may be prepared according to methods known generally in the art of expression of proteins using recombinant DNA technology.

25 2. Preparation of recombinant methionyl succinylated human leptin.

The present recombinant methionyl succinylated human leptin was prepared by reaction of
30 recombinant human leptin at ~150 mg/mL in sodium phosphate buffer at pH 7.0 with a 3-7 molar excess of succinic anhydride at 4°C for two hours. The reaction is quenched by addition of solid hydroxyl amine to a final concentration of 0.5M and pH 8.5. The reaction
35 mixture is then dialyzed vs. 10mM sodium phosphate,

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pH 7.0 and the mono-succinylated leptin form separated from di- and poly-succinylated leptin forms by ion exchange chromatography.

5 3. Preparation of zinc derivatized recombinant methionyl human leptin suspension.

 The present zinc derivatized recombinant methionyl human leptin suspension was prepared by taking a 752 μ l
10 sample of recombinant human leptin at 133 mg/mL in 1mM HCl and adding 48 μ l water, followed by 100 μ l of 500 μ M zinc chloride, followed by 100 μ l of 1M TRIS, pH 8.5. There is an immediate Zn:leptin precipitate that will rapidly fall out of solution at room
15 temperature, but which can be readily resuspended.

 4. Preparation of zinc derivatized recombinant methionyl human leptin lyophilized powder.

20

 Lyophilization of the present zinc derivatized recombinant methionyl human leptin lyophilized powder was carried out in a Virtis shelf lyophilizer. In short, a Zn:leptin suspension was
25 frozen on shelf at -50°C and held for 2 hours. The samples were annealed at -25°C for 2 hours and then frozen again at -50°C. The chamber pressure was lowered to 100 mTorr while maintaining shelf temperature at -50°C for 2 hours. Primary drying was
30 conducted by raising the shelf temperature to -25°C and holding for 10 hours while keeping the chamber pressure at 100mTorr. Secondary drying was conducted by ramping the shelf temperature to 25°C over 7 hours and holding for 10 hours while maintaining the chamber pressure at

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100mTorr. The cycle was ended at the end of secondary drying, the chamber was aerated to atmospheric pressure and lyophilized product removed from the lyophilizer.

5 5. Preparation of spray dried recombinant IL-1ra.

 The present spray dried recombinant IL-1ra protein was prepared by spray drying a 20 mg/mL IL-1ra solution, using a Buchi 190 mini spray dryer. The
10 inlet and outlet temperatures during spray drying were 130°C and 90°C, respectively. Feeding rate was 1-2 mL/min, and a spray-dried IL-1ra powder obtained.

 While the present invention has been
15 described in terms of certain preferred embodiments, it is understood that variations and modifications will occur to those skilled in the art. Therefore, it is intended that the appended claims cover all such equivalent variations which come within the scope of
20 the invention as claimed.

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CLAIMS:

1. A method for making a composition comprising an active ingredient contained within polymeric microparticles, wherein a mixture of the active ingredient and the polymer are dispersed within a continuous phase, the resulting dispersion is frozen, and the water and organic solvents removed from the dispersion by lyophilization.
2. A method according to Claim 1 wherein said continuous phase is aqueous or organic or a mixture thereof.
3. A method according to Claim 1 wherein the active ingredient-polymer mixture is obtained by dispersing an aqueous solution of the active ingredient in a second, non-aqueous phase containing the polymer, prior to addition to the continuous phase.
4. A method according to Claim 1 wherein the active ingredient-polymer mixture is obtained by dissolving both components in a non-aqueous solvent prior to addition to the continuous phase.
5. A method according to Claim 1 wherein the active ingredient is present as a dispersion of solid particles in a non-aqueous solution of the polymer, which is then added to the continuous phase.
6. A method according to Claim 1, wherein said active ingredient is omitted from said mixture, thereby producing blank polymeric microparticles.

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7. A method according to Claim 6 further comprising active ingredient being loaded onto said blank polymeric microparticles by suspending said blank polymeric microparticles in active ingredient solution.

5

8. A method according to any of Claims 1-7 wherein one or more excipients is combined with the active ingredient-polymer mixture prior to incorporation into the dispersion.

10

9. A method according to any of Claims 1-7 wherein said continuous phase contains one or more excipients.

15

10. A method according to any of Claims 1-7 wherein one or more excipients is mixed with the active ingredient and the same or a different excipient is present in the continuous phase.

20

11. A method according to Claim 1 wherein said polymer is selected from the group consisting of biodegradable and/or biocompatible polymers.

25

12. A method according to Claim 11 wherein said biodegradable polymers is selected from the group consisting of poly(lactide)s, poly(glycolide)s, poly(lactic acid)s, poly(glycolic acid)s, polyanhydrides, polyorthoesters, polyetheresters, polycaprolactone, polyesteramides, polycarbonate, polycyanoacrylate, polyurethanes, polyacrylate, blends and copolymers thereof.

30

13. A method according to Claim 12 wherein said polymer is poly(lactide-co-glycolide) (PLGA), and

- 30 -

wherein said polymer is dissolved in an organic solvent selected from the group consisting of chloroform, ethyl acetate, methylene chloride, acetonitrile, THF and acetone.

5

14. A method according to any of Claims 1-10 wherein the active ingredient is selected from the group consisting of peptides, small molecules, sugars, carbohydrates, nucleic acids, lipids, and proteins.

10

15. A method according to claim 14 wherein said active ingredient is a protein selected from the group consisting of G-CSF, GM-CSF, M-CSF, MGDF, the interferons (alpha, beta, and gamma), interferon consensus, the interleukins (1-12), erythropoietin (EPO), fibroblast growth factor, TNF, TNFbp, IL-1ra, stem cell factor, nerve growth factor, GDNF, BDNF, NT3, platelet-derived growth factor, tumor growth factor (alpha, beta), OPG, and leptin; or derivatives, analogs, fusions, conjugates, or chemically modified forms thereof.

20

16. A method according to Claim 15 wherein said protein is leptin, or a derivative, analog, fusion, conjugate, or chemically modified form thereof.

25

17. A method according to Claim 16 wherein said modified form of leptin is selected from the group consisting of Fc-leptin fusion, succinylated-leptin, and zinc derivatized leptin.

30

18. A method according to Claim 15 wherein said protein is G-CSF, or a derivative, analog, fusion, conjugate, or chemically modified form thereof.

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19. A method according to Claim 15 wherein said protein is BDNF, or a derivative, analog, fusion, conjugate, or chemically modified form thereof.

5

20. A pharmaceutical composition for the sustained-release of an active ingredient, said composition produced by the method of any of Claims 1-15.

10

21. A pharmaceutical composition comprising leptin, or a derivative, analog, fusion, conjugate, or chemically modified form thereof contained within a polymeric microparticle.

15

22. A pharmaceutical composition comprising G-CSF, or a derivative, analog, fusion, conjugate, or chemically modified form thereof contained within a polymeric microparticle.

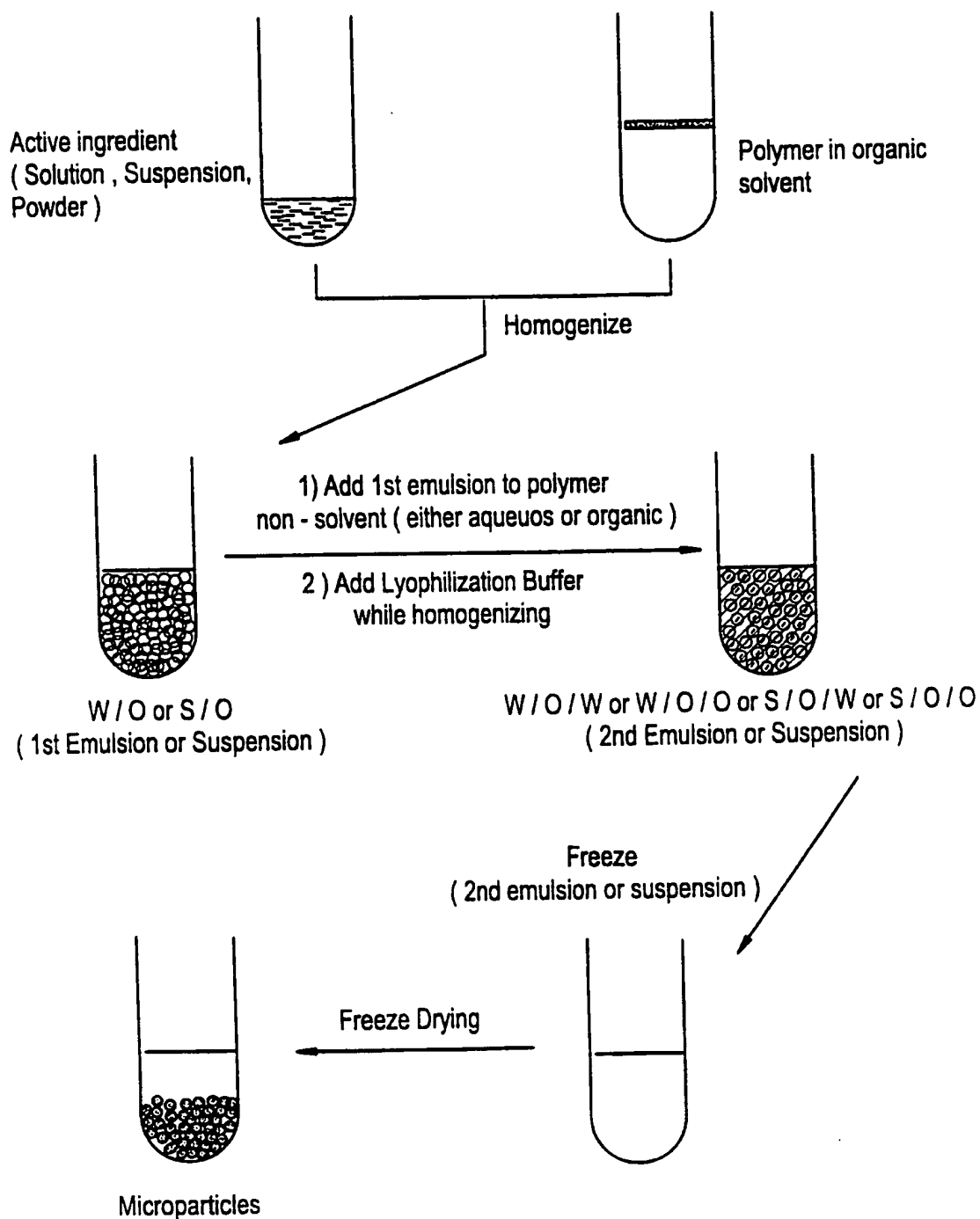
20

23. A pharmaceutical composition comprising BDNF, or a derivative, analog, fusion, conjugate, or chemically modified form thereof contained within a polymeric microparticle.

25

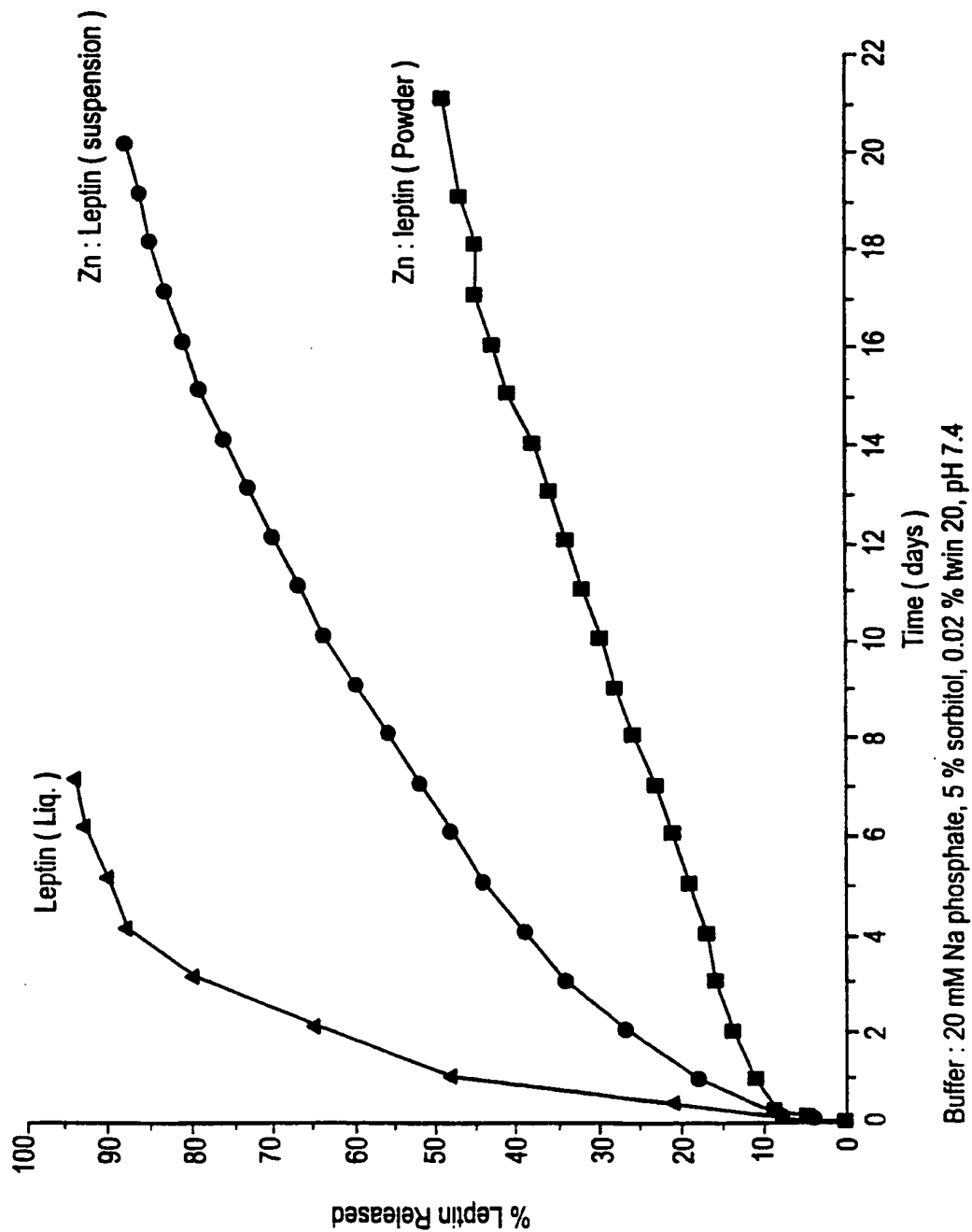
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FIG.1



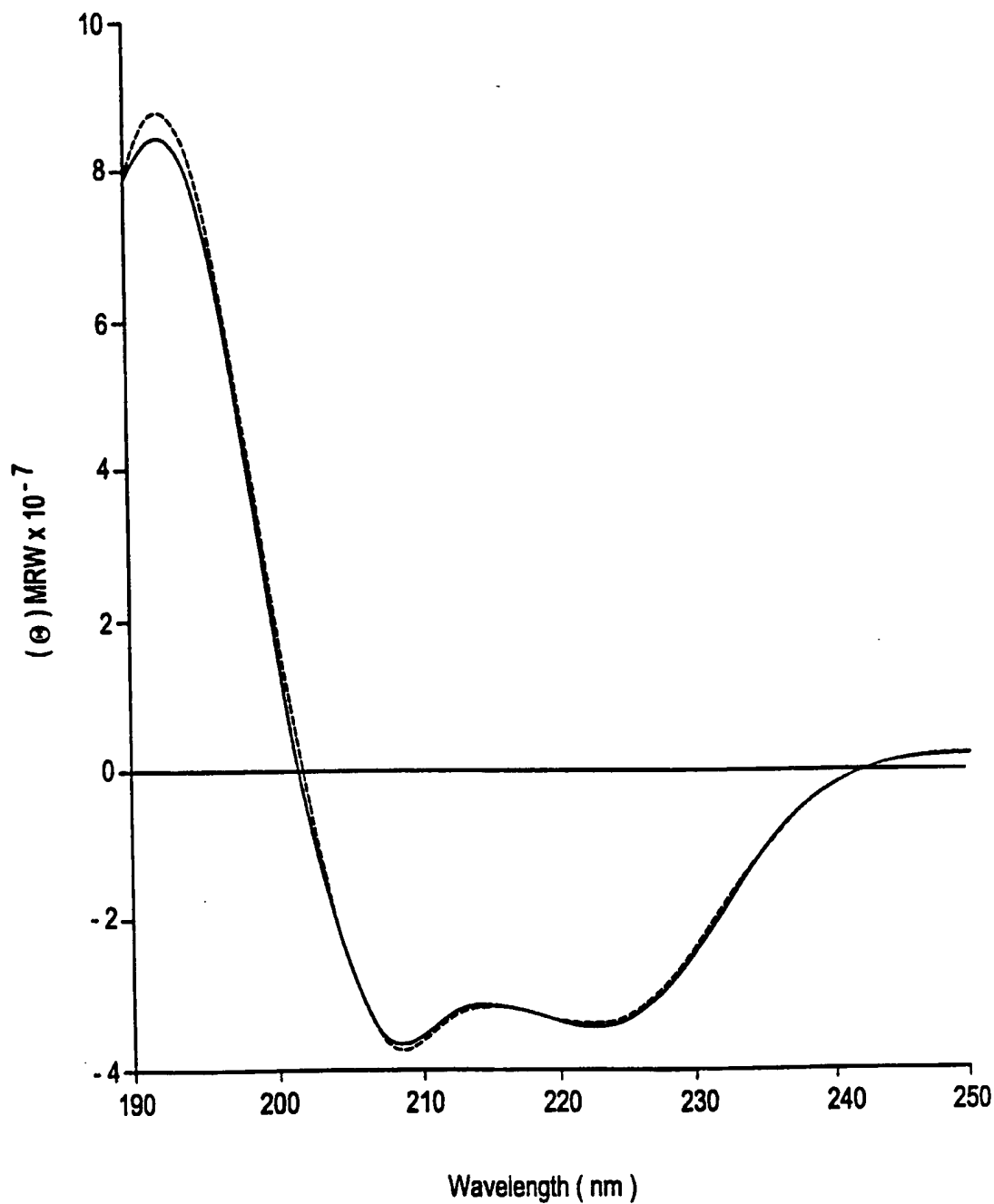
2 / 7

FIG.2



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FIG.3

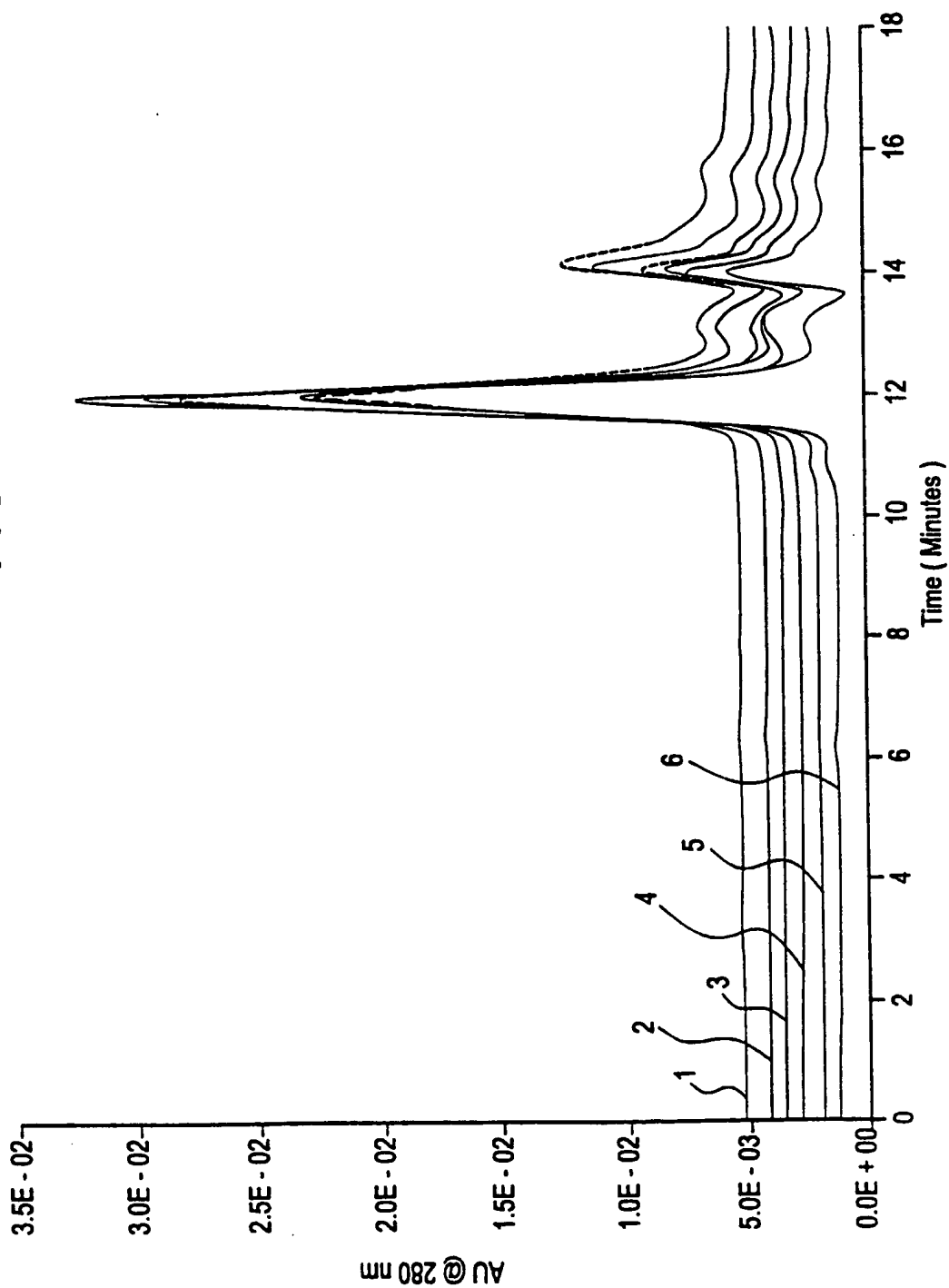


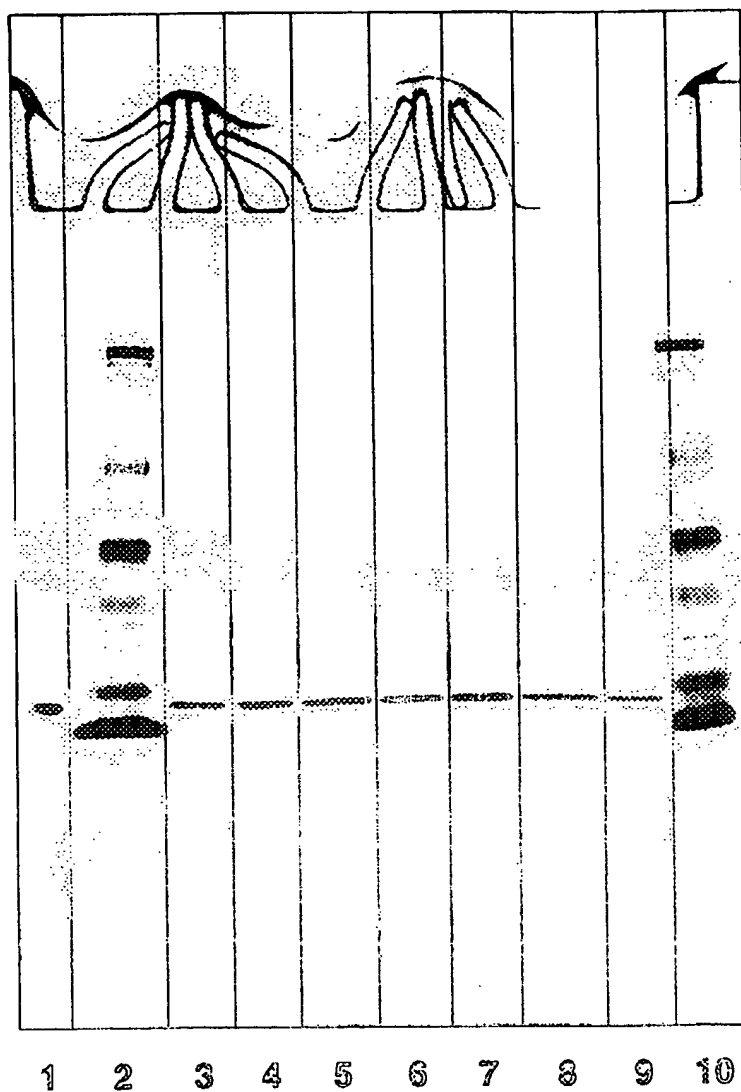
----- Leptin in Formulation Buffer

———— Leptin from Microparticles on seventh day of in vitro release

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FIG. 4



5/7
FIG. 5

Lane #	Sample Name
1	Leptin Standard
2	Mol. Weight Standards
3	2 hrs Release Sample
4	24 hrs Release Sample
5	68 hrs Release Sample
6	92 hrs Release Sample
7	116 hrs Release Sample
8	140 hrs Release Sample
9	168 hrs Release Sample
10	Mol. Weight Standards

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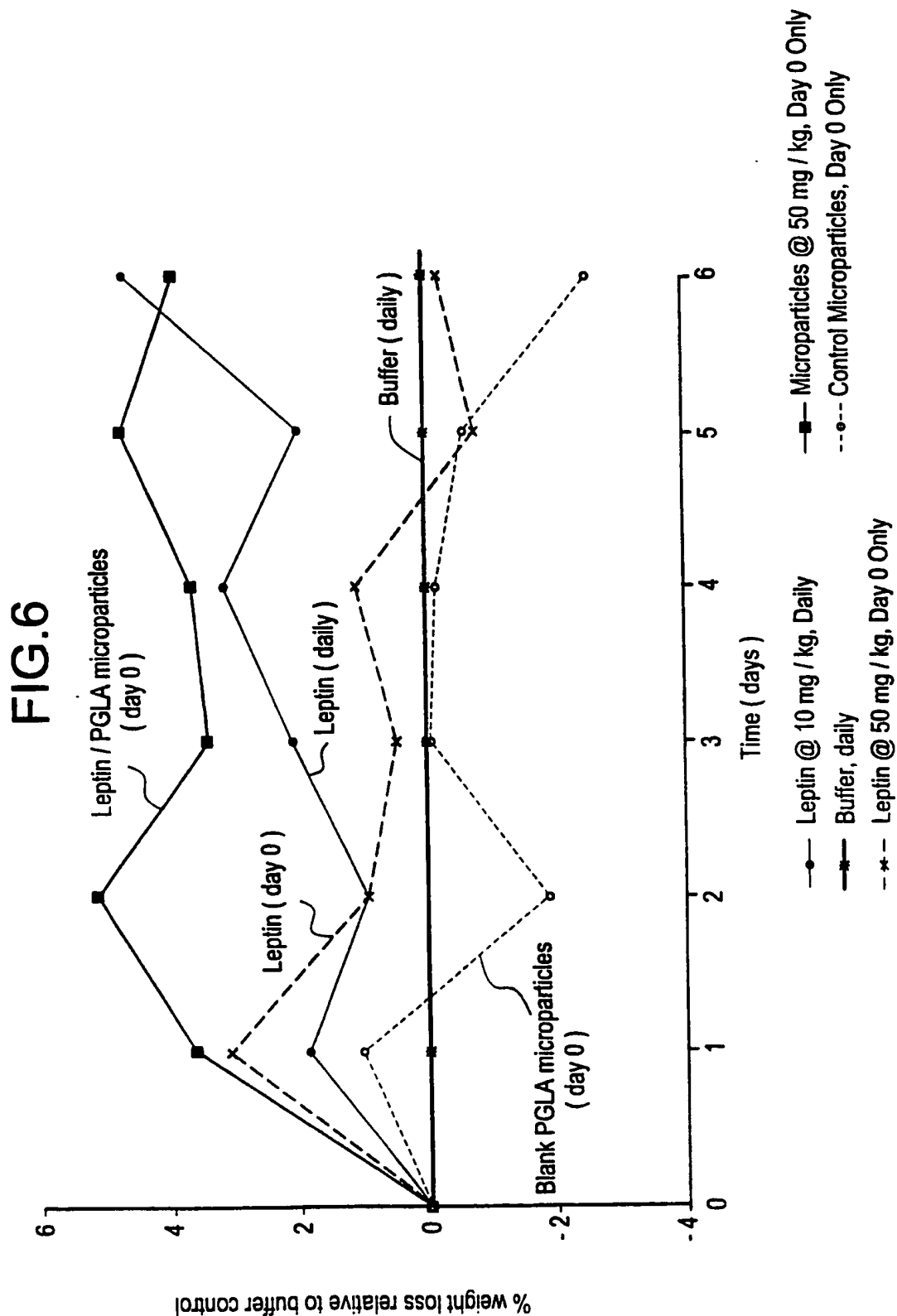
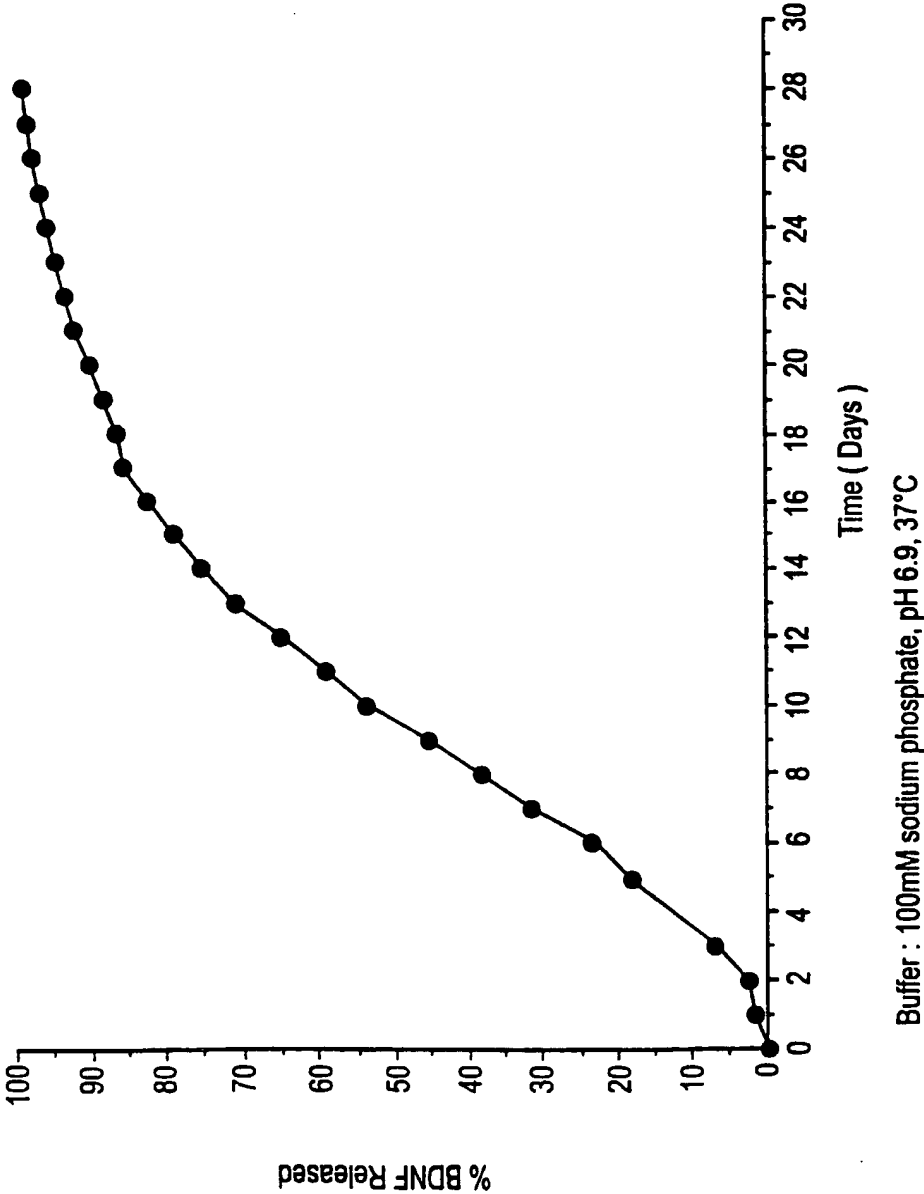


FIG.7



INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 98/07856

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 A61K9/16

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	EP 0 140 085 A (FUJISAWA PHARMACEUTICAL) 8 May 1985 see the whole document ---	1-23
A	H. T. WANG: "Influence of formulation methods on the in vitro controlled release of protein from poly(ester) microspheres" JOURNAL OF CONTROLLED RELEASE, no. 17, 1991, pages 21-23, XP000223264 see the whole document ---	1-23
A	WO 96 22786 A (TAKEDA CHEMICAL INDUSTRIES) 1 August 1996 see the whole document ---	1-23
-/--		

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

16 July 1998

Date of mailing of the international search report

23/07/1998

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INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 98/07856

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>EP 0 442 671 A (TAKEDA CHEMICAL INDUSTRIES) 21 August 1991 see the whole document</p>	1-23

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

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